

---

# AKADEMİK PERSPEKTİFTEN SAĞLIK BİLİMLERİ

Editör: Dr.Öğr.Üyesi Necati BÜKECİK

---



**yaz**  
yayınları

# **Akademik Perspektiften Saęlık Bilimleri**

**Editör**

Dr.Öęr.Üyesi Necati BÜKECİK

**yaz**  
yayınları

2025

## **Akademik Perspektiften Sağlık Bilimleri**

Editör: Dr.Öğr.Üyesi Necati BÜKECİK

---

### **© YAZ Yayınları**

Bu kitabın her türlü yayın hakkı Yaz Yayınları'na aittir, tüm hakları saklıdır. Kitabın tamamı ya da bir kısmı 5846 sayılı Kanun'un hükümlerine göre, kitabı yayınlayan firmanın önceden izni alınmaksızın elektronik, mekanik, fotokopi ya da herhangi bir kayıt sistemiyle çoğaltılamaz, yayınlanamaz, depolanamaz.

---

E\_ISBN 978-625-5838-37-7

Haziran 2025 – Afyonkarahisar

Dizgi/Mizanpaj: YAZ Yayınları

Kapak Tasarım: YAZ Yayınları

YAZ Yayınları. Yayıncı Sertifika No: 73086

M.İhtisas OSB Mah. 4A Cad. No:3/3  
İscehisar/AFYONKARAHİSAR

[www.yazyayinlari.com](http://www.yazyayinlari.com)

[yazyayinlari@gmail.com](mailto:yazyayinlari@gmail.com)

[info@yazyayinlari.com](mailto:info@yazyayinlari.com)

## İÇİNDEKİLER

- Innovative Nursing Programs to Improve Public Health: A Review on Telemedicine and Chronic Disease Management.....1**  
*Necati BÜKECİK, Filiz ADANA*
- Görme Yetersizliği Olan Çocuklara Yönelik Günlük Yaşam Aktiviteleri (GYA) Becerilerine Yönelik Yardımcı Yöntemler .....18**  
*Çiğdem Müge HAYLI, Ramazan KARATAŞ*
- Mitophagy Mechanism and Its Relationship With Diseases.....29**  
*Canan CEYLAN, Nurcan DÖNMEZ*
- Kedi ve Köpeklerde Gastrointestinal Yabancı Cisimlerin Cerrahi Olarak Değerlendirmesi.....53**  
*Bahar ERDEN, Mustafa Barış AKGÜL, Sevdet KILIÇ*
- TİM-3: Kanser İmmünoterapisinde Potansiyel Bir Bağışıklık Kontrol Noktası .....71**  
*Zeynep Burcu MEKİK, Altınay ALTINKAYNAK, Gizem TUTKUN, Ece ŞİMŞEK, Kübra YILDIRIM, Emel KAYNAKÇI, Ahmet Yılmaz ÇOBAN*
- Anatomical Variations and Pathologies of The Maxillary Sinus in Dentistry Practice .....84**  
*Emine ARARAT*
- RNAi Mechanism-Mediated Gene Suppression and Its Therapeutic Efficacy .....100**  
*Muhammet Mücahit SARI, Ayşe AYDOĞAN*

**Kadınlarda Sağlık Eşitsizliği ve Sağlığın  
Geliştirilmesinde İnternet Kullanımı .....127**  
*Seda GÜRAY, Zeynep Seyyide KAYA*

**Akut Görme Kaybı Nedenleri: Acil Tanı ve Tedavi  
Yaklaşımları.....140**  
*Aydın MAÇİN*

**Web of Science Veri Tabanındaki Deneysel Epilepsi Konulu  
Yayımların Bibliyometrik Analizi .....161**  
*Ramazan GÜNEŞER*

*"Bu kitapta yer alan bölümlerde kullanılan kaynakların, görüşlerin, bulguların, sonuçların, tablo, şekil, resim ve her türlü içeriğin sorumluluğu yazar veya yazarlarına ait olup ulusal ve uluslararası telif haklarına konu olabilecek mali ve hukuki sorumluluk da yazarlara aittir."*

# **MITOPHAGY MECHANISM AND ITS RELATIONSHIP WITH DISEASES**

**Canan CEYLAN<sup>1</sup>**

**Nurcan DÖNMEZ<sup>2</sup>**

## **1. INTRODUCTION**

Mitochondria, essential organelles that serve as the cellular energy centers in all eukaryotic cells, are responsible for aerobic respiration and the production of adenosine triphosphate (ATP) through oxidative phosphorylation (Nunnari & Suomalainen, 2012; Özsan, 2023). In addition to being the primary organelles for reactive oxygen species (ROS) production during the electron transport chain, mitochondria are also exposed to oxidative stress under conditions of structural and functional impairment (Özsan, 2023; Wong & Holzbaur, 2014).

Mitophagy functions as a specific and selective type of autophagy. It involves the transport and degradation of damaged cytotoxic mitochondria into lysosomes via a selective autophagic mechanism. At the same time, it preserves mitochondrial structural and functional integrity as well as intracellular homeostasis by maintaining the balance of mitochondrial quality and quantity necessary for normal cell physiology and tissue development (Bravo-San Pedro et al., 2017). This process plays a critical role in maintaining

---

<sup>1</sup> Dr. Öğr. Üyesi, İstanbul Sabahattin Zaim Üniversitesi, Spor Bilimleri Fakültesi, Beden Eğitimi ve Spor Öğretmenliği, canan.ceylan@izu.edu.tr, ORCID: 0000-0002-0424-4403.

<sup>2</sup> Prof. Dr., Selçuk Üniversitesi, Veteriner Fakültesi, Fizyoloji ABD, nurcandonmez@selcuk.edu.tr, ORCID: 0000-0003-4271-598X.

mitochondrial homeostasis, ensuring cellular health, and regulating various physiological processes. Mitophagy is activated by stressors such as nutrient deficiency, hypoxia, DNA damage, inflammation, and mitochondrial membrane depolarization that disrupt mitochondrial homeostasis. However, the failure to eliminate defective mitochondria and the suppression of the mitophagy mechanism may lead to increased ROS production, inflammation, and apoptosis, resulting in a wide range of pathological outcomes such as diabetes mellitus, neurodegenerative diseases, and cancer. Recent studies have shown that impaired mitophagy is a significant factor in the pathogenesis of many human diseases, including neurodegenerative disorders, cancer, cardiovascular diseases, and metabolic disorders (Palikaras et al., 2017).

This review aims to provide a general overview of the regulatory effects of mitophagy-associated factors and proteins, focusing on their roles in cellular homeostasis, physiological function, and disease development.

## **2. MITOCHONDRIAL DYNAMICS**

Fusion and fission are fundamental processes that govern mitochondrial dynamics, ensuring proper mitochondrial shape, distribution, and function. These dynamic events are controlled by guanosine triphosphatases (GTPases), particularly those belonging to the dynamin superfamily. Dynamin-related protein 1 (DRP1) is a cytosolic GTPase that translocates to mitochondria to facilitate fission. In contrast, the mitofusin proteins MFN1 and MFN2 are crucial mediators of outer mitochondrial membrane fusion. OPA1, another GTPase situated within the inner mitochondrial membrane and intermembrane space, is essential for modulating both the fusion and fission of the inner membrane. Notably, mutations in OPA1,

along with heightened cellular vulnerability to apoptosis, have been associated with the development of optic atrophy (Cipolat et al., 2004; Ferguson & De Camilli, 2012; Youle & Van Der Blik, 2012).

Healthy mitochondria contribute to the exchange of genetic material and the preservation of mitochondrial morphology and function through the fusion and fission mechanisms involved in mitochondrial dynamics. The dynamics of mitochondrial fusion and fission are regulated by DRP1, MFN1, MFN2, and OPA1 proteins. Mitochondrial fission contributes to mitochondrial apoptosis and is necessary for the initiation of mitophagy, whereas mitochondrial fusion has an enhancing effect on mitochondrial metabolism. Under cellular stress, damaged mitochondria undergo fission from the mitochondrial network or are isolated due to DRP1 activation or mitochondrial membrane potential dysfunction in the autophagy system (Montava-Garriga & Ganley, 2020). As a result of mitochondrial imbalance, fusion and fission processes are disrupted, and the mitophagy mechanism is activated to degrade fragmented mitochondria (Shirihai et al., 2015). Consequently, the isolated mitochondria can be cleared through different mitophagy pathways involving both ubiquitin-dependent and ubiquitin-independent mitophagy receptors.

### **3. MITOPHAGY MECHANISM**

The most prominent mechanism of mitochondrial turnover is autophagy. Defined in Greek as “self-eating,” autophagy is a catabolic process that allows eukaryotic cells to degrade cytoplasmic components, recover vital metabolites, and protect against toxicity caused by damaged organelles or harmful aggregates. Three main types of autophagy have been

identified in eukaryotic cells: macroautophagy, chaperone-mediated autophagy, and microautophagy (Boya et al., 2013).

One of the macroautophagy subtypes, mitochondrial autophagy (mitophagy), was first described by Lemasters in 2005 as a selective intracellular degradation mechanism targeting dysfunctional mitochondria. Mitophagy plays an important role in maintaining the homeostasis of the intracellular mitochondrial quality control system and regulating mitochondrial energy metabolism (Ajoolabady et al., 2022). Cells support mitophagy mechanisms through various stimuli and signaling cascades with the help of numerous regulators. In the regulation of mitophagy, both ubiquitin-dependent and ubiquitin-independent pathways operate (Khaminets et al., 2016).

### **3.1. Ubiquitin-Dependent Pathways**

#### **3.1.1. PINK1-Parkin Mediated Mitophagy**

The ubiquitin-dependent pathway supports complex processes involving proteins anchored to the mitochondrial surface in order to promote mitophagy. The PTEN-induced putative kinase 1 (PINK1)-Parkin pathway acts as a sensor that detects cellular stress and removes damaged mitochondria, serving as a key regulator of ubiquitin-dependent mitophagy (Sekine & Youle, 2018).

Under normal physiological conditions, PINK1 is imported from the cytosol into the inner mitochondrial membrane by interacting with the translocase complex located on the outer mitochondrial membrane. Within the intermembrane space, it undergoes proteolytic cleavage by the presenilin-associated rhomboid-like protease (PARL), after which the truncated PINK1 is released back into the cytosol and targeted for degradation via the ubiquitin-proteasome system (UPS) (Vives-Bauza et al., 2010; Yamano & Youle, 2013). In

contrast, when mitochondria experience depolarization and lose their membrane potential, this import and cleavage process is disrupted. Consequently, PINK1 accumulates on the outer mitochondrial membrane instead of reaching the inner membrane. This accumulation acts as a signal, recruiting the cytosolic E3 ubiquitin ligase Parkin to the mitochondria in a PINK1-dependent manner (Fiesel et al., 2023). These sequential molecular events facilitate the delivery of dysfunctional mitochondria to autophagosomes, where they are subsequently degraded in autolysosomes, thereby concluding the mitophagy process (Uoselis et al., 2023). The PINK1–Parkin pathway mediates mitochondrial quality control mechanisms such as mitochondrial-derived vesicles and mitochondrial dynamics to maintain energy homeostasis. PINK1 indirectly stimulates dynamin-related protein 1 (DRP1) activation, inducing the fission of damaged Mitochondria (Pryde et al., 2016). On the other hand, Parkin-dependent degradation of abnormal and misfolded mitofusins (MFNs) disrupts mitochondrial fusion, thereby isolating defective organelles from the healthy mitochondrial network and preserving mitochondrial homeostasis (Tanaka et al., 2010). Additionally, PINK1 phosphorylates MFN2 and triggers mitochondrial separation, promoting the removal of damaged organelles in cooperation with Parkin (Y. Chen & Dorn, 2013).

### **3.2. Ubiquitin-Independent Pathways**

Mitophagy is induced by various cellular stress signals and structural changes. Dysfunction of mitochondrial membrane potential is a strong stimulus for mitophagy (Elmore et al., 2001). Unlike the PINK1/Parkin pathway, mitophagy receptors primarily contribute to the remodeling of the mitochondrial network in response to peripheral stress, cellular reprogramming, or differentiation (J. Zhang & Ney, 2009).

Ubiquitin-independent mitophagy is primarily regulated by two major classes of receptors that facilitate mitochondrial clearance under both normal and stress conditions. One group comprises BCL2/adenovirus E1B 19 kDa-interacting protein 3 (BNIP3) and its homolog NIX, while the other includes FUN14 domain-containing protein 1 (FUNDC1). NIX, an outer mitochondrial membrane protein, plays a pivotal role in the targeted elimination of dysfunctional mitochondria, particularly during the maturation of reticulocytes when mitochondria are removed from developing erythrocytes (Church & Margolis, 2024). Both BNIP3 and NIX are known to enhance mitophagy in response to hypoxic conditions through the stabilization of hypoxia-inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ), which promotes mitochondrial turnover (Jung et al., 2019). Under hypoxic stress, BNIP3 expression increases and localizes to the outer mitochondrial membrane via its C-terminal transmembrane domain. Mutations in BNIP3 that impair its ability to homodimerize—without affecting mitochondrial targeting—lead to mitophagy dysfunction, highlighting its essential role as an initiator of mitochondrial degradation (Hanna et al., 2012; Zhu et al., 2013). As with other mitophagy receptors, BNIP3 harbors an LC3-interacting region (LIR) motif in its N-terminal segment. This motif enables direct binding to LC3, independent of ubiquitin tagging, thus allowing the damaged mitochondria to be sequestered into autophagosomes for selective degradation via the autophagic pathway (Bai et al., 2023).

FUNDC1, another outer mitochondrial membrane protein, functions as a receptor for hypoxia-related mitophagy (Liu et al., 2012). Like BNIP3, FUNDC1 supports mitochondrial fission in response to stress by recruiting DRP1 to mitochondria (Chen et al., 2016; Landes et al., 2010).

## **4. THE EFFECT OF MITOPHAGY ON DISEASES**

### **4.1. Metabolic Diseases**

Chronic hyperglycemia and insulin resistance, which are among the major symptoms of type 2 diabetes mellitus (T2DM), are associated with mitochondrial damage (Özsan & Ceylan, 2023). High glucose concentrations lead to mitochondrial dysfunction by increasing oxidative stress, which results in elevated mitochondrial ROS production and tissue damage (Rovira-Llopis et al., 2017). Therefore, the preservation of mitochondrial quality through mitophagy is vital for preventing disease progression.

Persistently high glucose levels promote mitochondrial fission, which ensures mitochondrial dynamics, while decreasing mitochondrial fusion due to DRP1 recruitment and impairment of OPA1/MFN function (Rovira-Llopis et al., 2017). Compared to healthy controls, mitophagy flux is reported to be impaired in T2DM patients, whose mitochondria are smaller in size (Bhansali et al., 2017). Expressions of mitophagy-related proteins, including NIX, PINK1, and Parkin, have been shown to increase in prediabetic individuals with mild hyperglycemia but decrease in patients with T2DM (Scheele et al., 2007). Thus, elevated ROS levels associated with T2DM may contribute to the suppression of mitophagy and the accumulation of dysfunctional mitochondria.

Obesity, which significantly increases the risk of developing T2DM, shares several common mechanisms involving mitophagy. In obesity, mitochondrial quality in adipose tissue is impaired, and disruption of the balance between mitochondrial fission and fusion leads to mitochondrial damage, resulting in a reduced mitophagic response (Greene et al., 2015).

## **4.2. Cardiovascular Diseases**

The heart muscle is a vital organ that primarily generates adenosine triphosphate (ATP) through oxidative phosphorylation. Mitochondria occupy approximately 30% of the cardiac muscle volume and produce about 6 kg of ATP daily to support the pumping power of the heart. This high demand of the heart is directly associated with the function of myocardial cells and mitochondrial quality control (Han et al., 2017; Sciarretta et al., 2018). Dysfunctional mitophagy correlates strongly with mitochondrial damage and reactive oxygen species (ROS) formation, both of which contribute to the pathogenesis of cardiovascular diseases (Lu et al., 2023). Dysfunctional mitochondria produce less ATP and excessive ROS. Increased sensitivity of cardiac muscle to oxidative stress and excessive ROS accumulation can lead to chronic damage processes such as increased ischemia/reperfusion injury (IRI), heart failure, apoptosis, and fibrosis, ultimately resulting in cardiac dysfunction and cardiomyocyte death (Whelan et al., 2010).

Studies have shown that dysfunctional mitophagy in cardiac tissue is associated with increased susceptibility to stress and the development of heart failure. Parkin expression in the heart increases rapidly in response to mitochondrial damage, suggesting an adaptive response of the heart to cellular stress. In experimental mice deficient in Parkin, an accumulation of dysfunctional mitochondria, increased ventricular remodeling, and progression to heart failure were observed after myocardial infarction (Han et al., 2017; Ikeda et al., 2015; Kubli et al., 2013). Indeed, alterations in mitophagy regulators in mouse models result in various forms of cardiac dysfunction. Furthermore, mice deficient in PINK1, BNIP3, NIX, and ATG5 develop various cardiac defects, including cardiomyopathy, cardiac hypertrophy, and accumulation of dysfunctional Mitochondria (Billia et al., 2011; Dorn, 2010).

### **4.3. Neurodegenerative Diseases**

Mitophagy, a mitochondrial quality control process, regulates mitochondrial function to preserve neuronal health (Dias et al., 2013; Rugarli & Langer, 2012). Mitochondrial damage and impaired mitophagy play key roles in the pathogenesis of many neurodegenerative diseases, including Parkinson's and Alzheimer's diseases (Özsan et al., 2025).

Parkinson's disease is a common neurodegenerative disorder characterized by the loss of dopaminergic neurons in the substantia nigra and the accumulation of mutated  $\alpha$ -synuclein aggregates (Dauer William & Przedborski Serge, 2003; Kısadere et al., 2021). PINK1 is highly expressed in organs and tissues with high energy demand, including the brain, heart, and muscles (Suen et al., 2010). Loss of PINK1 or Parkin activity impairs mitophagy by reducing the recruitment of ubiquitin-binding proteins and promoting excessive mitochondrial fusion through accumulation of MFN1 and MFN2 (Burté et al., 2015). Mutations in PINK1 and Parkin are involved in the pathogenesis of Parkinson's disease and suppress the formation of mitophagy (Pickrell & Youle, 2015).

Alzheimer's disease is an age-related neurodegenerative disorder characterized by neuronal cell death in the cerebral cortex, leading to severe memory loss and cognitive decline ( Kısadere et al., 2019; Özsan et al., 2025). Accumulation of damaged mitochondria, synaptic degeneration, amyloid- $\beta$  oligomers derived from amyloid precursor protein, and intracellular neurofibrillary tangles are all implicated in the pathogenesis of Alzheimer's disease (Cen et al., 2021). In Alzheimer's disease, the levels of DRP1 involved in mitochondrial fission and OPA1 involved in mitochondrial fusion are reported to be decreased. These changes lead to neuronal abnormalities, mitochondrial dysfunction, and

excessive accumulation of amyloid- $\beta$ , ultimately suppressing mitophagy and resulting in neuronal death (Li et al., 2016; Song et al., 2021).

#### **4.4. Cancer**

Cancer adversely affects human health through the abnormal proliferation and differentiation of local tissue cells caused by various factors that lead to genetic damage and tumor formation (Panigrahi et al., 2020a).

Mitophagy is a key regulator in determining whether a cell survives or undergoes programmed death. While its excessive activation may result in mitochondrial depletion, leading to inadequate ATP production and eventual cell demise, a balanced mitophagic response is crucial for maintaining intracellular homeostasis and promoting cell viability. In the context of malignant neoplasms, mitophagy has been shown to contribute to the dysregulated growth and persistence of cancer cells, functioning in a dual capacity as both a promoter of tumorigenesis and a potential tumor suppressor (Chang et al., 2017a). The interplay between these opposing roles is pivotal in influencing whether a tumor progresses or regresses through apoptotic mechanisms.

Mitophagy appears to be functionally important in tumor suppression through the removal of damaged mitochondria that may contribute to carcinogenesis (Vara-Perez et al., 2019). Lauren et al. (2017) reported that mitophagy inhibits tumor growth by eliminating damaged mitochondria; otherwise, dysfunctional mitochondria alter cell structure and may directly or indirectly trigger tumor formation (Drake et al., 2017). Accordingly, different levels of mitophagy have been observed in various cancers, including rectal, lung, and breast cancer, compared to normal conditions (Ferro et al., 2020).

Although cell proliferation is associated with metabolic reprogramming that inhibits oxidative phosphorylation and enhances glycolytic flux, cancer cells typically exhibit decreased oxidative phosphorylation and increased glycolysis. During cancer progression, some cancer cells shift mitochondrial aerobic respiration to glycolysis to meet their energy demands—a phenomenon known as the Warburg effect. The Warburg effect is characterized by abnormal mitochondrial quality control, increased ROS production, altered redox regulation, and deficient apoptotic signaling (Panigrahi et al., 2020b). In the mitophagy process, Parkin suppresses the Warburg effect by enhancing oxidative metabolism (Zhang et al., 2011). Therefore, Parkin can be considered to exhibit tumor-suppressive activity.

The PINK1/Parkin pathway serves as a central mediator of mitophagy. Parkin dysfunction can lead to mitophagy inhibition and promote carcinogenesis in various cancer types. Mutations in these two proteins have been detected in numerous tumors, including lung cancer, ovarian cancer, glioblastoma/glioma, colorectal cancer, and breast cancer (Chang et al., 2017b). Inhibition of mitophagy during cancer development can significantly impair the function of cellular systems (cells, tissues, organs) and lead to increased ROS levels (Fujiwara et al., 2008; Youle & Narendra, 2011). Indeed, elevated ROS can cause DNA damage and promote gene expression dysfunction, ultimately contributing to cancer progression (Um & Yun, 2017).

When exposed to hypoxia or cellular stress, mitochondria undergo a decline in membrane potential, initiating the autophagic clearance of dysfunctional organelles. This depolarization mechanism plays a significant role in shaping the malignant characteristics of tumor cells (Ferro et al., 2020; Panigrahi et al., 2020c). In this context, the outer mitochondrial membrane proteins BNIP3 and its paralog NIX

act as pivotal regulators of mitophagy activation under hypoxic conditions. Hypoxia-inducible factor 1 (HIF-1) enhances the expression of both NIX and BNIP3 in tumor environments, and NIX-driven mitophagy has been implicated in the progression of glioblastoma (Sowter et al., 2001). Notably, suppression of the NIX receptor disrupts the survival of cancer stem cells by promoting reactive oxygen species (ROS) clearance and triggering apoptosis in tumor populations (Jung et al., 2019).

## **5. CONCLUSION**

The multifaceted role of mitochondria in cellular energy production, metabolic regulation, and redox balance has attracted increasing scientific interest, particularly in the context of how these functions intersect with the maintenance of cellular homeostasis and the regulation of mitophagy under pathological conditions. The recognition and degradation of damaged mitochondria through autophagosomes are achieved via the coordinated action of various mitophagic mechanisms. Mitophagy is a homeostatic process that plays a critical role in both the preservation of cellular health and the pathogenesis of numerous diseases. Functional impairments in different mitophagy pathways directly contribute to the onset and progression of various disorders. Therefore, comprehensive research is required to understand how diseases are influenced by mitophagy and to identify potential mitophagy-regulating factors that play a central role in these processes.

## REFERENCES

- Ajoolabady, A., Chiong, M., Lavandero, S., Klionsky, D. J., & Ren, J. (2022). Mitophagy in cardiovascular diseases: molecular mechanisms, pathogenesis, and treatment. In *Trends in Molecular Medicine* (Vol. 28, Issue 10, pp. 836–849). Elsevier Ltd. <https://doi.org/10.1016/j.molmed.2022.06.007>
- Bai, H., Fang, Y., Cao, H., Xing, C., Zhang, C., Zhuang, Y., Guo, X., Li, G., Hu, M., Hu, G., & Yang, F. (2023). Inhibition of the BNIP3/NIX-dependent mitophagy aggravates copper-induced mitochondrial dysfunction in duck renal tubular epithelial cells. *Environmental Toxicology*, 38(3), 579–590. <https://doi.org/10.1002/tox.23704>
- Bhansali, S., Bhansali, A., Walia, R., Saikia, U. N., & Dhawan, V. (2017). Alterations in mitochondrial oxidative stress and mitophagy in subjects with prediabetes and type 2 diabetes mellitus. *Frontiers in Endocrinology*, 8(DEC). <https://doi.org/10.3389/fendo.2017.00347>
- Billia, F., Hauck, L., Konecny, F., Rao, V., Shen, J., & Mak, T. W. (2011). PTEN-inducible kinase 1 (PINK1)/Park6 is indispensable for normal heart function. *Proceedings of the National Academy of Sciences of the United States of America*, 108(23), 9572–9577. <https://doi.org/10.1073/pnas.1106291108>
- Boya, P., Reggiori, F., & Codogno, P. (2013). *Emerging regulation and functions of autophagy*.
- Bravo-San Pedro, J. M., Kroemer, G., & Galluzzi, L. (2017). Autophagy and Mitophagy in Cardiovascular Disease. In *Circulation Research* (Vol. 120, Issue 11, pp. 1812–1824). Lippincott Williams and Wilkins. <https://doi.org/10.1161/CIRCRESAHA.117.311082>

- Burté, F., Carelli, V., Chinnery, P. F., & Yu-Wai-Man, P. (2015). Disturbed mitochondrial dynamics and neurodegenerative disorders. In *Nature Reviews Neurology* (Vol. 11, Issue 1, pp. 11–24). Nature Publishing Group.  
<https://doi.org/10.1038/nrneurol.2014.228>
- Cen, X., Zhang, M., Zhou, M., Ye, L., & Xia, H. (2021). Mitophagy regulates neurodegenerative diseases. In *Cells* (Vol. 10, Issue 8). MDPI.  
<https://doi.org/10.3390/cells10081876>
- Chang, J. Y., Yi, H. S., Kim, H. W., & Shong, M. (2017a). Dysregulation of mitophagy in carcinogenesis and tumor progression. In *Biochimica et Biophysica Acta - Bioenergetics* (Vol. 1858, Issue 8, pp. 633–640). Elsevier B.V.  
<https://doi.org/10.1016/j.bbabi.2016.12.008>
- Chang, J. Y., Yi, H. S., Kim, H. W., & Shong, M. (2017b). Dysregulation of mitophagy in carcinogenesis and tumor progression. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, 1858(8), 633–640.  
<https://doi.org/10.1016/J.BBABIO.2016.12.008>
- Chen, M., Chen, Z., Wang, Y., Tan, Z., Zhu, C., Li, Y., Han, Z., Chen, L., Gao, R., Liu, L., & Chen, Q. (2016). Mitophagy receptor FUNDC1 regulates mitochondrial dynamics and mitophagy. *Autophagy*, 12(4), 689–702.  
<https://doi.org/10.1080/15548627.2016.1151580>
- Chen, Y., & Dorn, G. W. (2013). PINK1-phosphorylated mitofusin 2 is a parkin receptor for culling damaged mitochondria. *Science*, 340(6131), 471–475.  
<https://doi.org/10.1126/science.1231031>

- Church, T. R., & Margolis, S. S. (2024). Mechanisms of ubiquitin-independent proteasomal degradation and their roles in age-related neurodegenerative disease. In *Frontiers in Cell and Developmental Biology* (Vol. 12). Frontiers Media SA. <https://doi.org/10.3389/fcell.2024.1531797>
- Cipolat, S., Martins De Brito, O., Zilio, B. D., & Scorrano, L. (2004). *OPA1 requires mitofusin 1 to promote mitochondrial fusion*. <https://www.pnas.org>
- Dauer William, & Przedborski Serge. (2003). Parkinson's Disease: Review Mechanisms and Models. *Neuron*, 39, 889–909.
- Dias, V., Junn, E., & Mouradian, M. M. (2013). The role of oxidative stress in parkinson's disease. In *Journal of Parkinson's Disease* (Vol. 3, Issue 4, pp. 461–491). I O S Press. <https://doi.org/10.3233/JPD-130230>
- Dorn, G. W. (2010). Mitochondrial pruning by nix and BNip3: An essential function for cardiac-expressed death factors. In *Journal of Cardiovascular Translational Research* (Vol. 3, Issue 4, pp. 374–383). <https://doi.org/10.1007/s12265-010-9174-x>
- Drake, L. E., Springer, M. Z., Poole, L. P., Kim, C. J., & Macleod, K. F. (2017). Expanding perspectives on the significance of mitophagy in cancer. In *Seminars in Cancer Biology* (Vol. 47, pp. 110–124). Academic Press. <https://doi.org/10.1016/j.semcancer.2017.04.008>
- Ferguson, S. M., & De Camilli, P. (2012). Dynamin, a membrane-remodelling GTPase. In *Nature Reviews Molecular Cell Biology* (Vol. 13, Issue 2, pp. 75–88). <https://doi.org/10.1038/nrm3266>

- Ferro, F., Servais, S., Besson, P., Roger, S., Dumas, J. F., & Brisson, L. (2020). Autophagy and mitophagy in cancer metabolic remodelling. In *Seminars in Cell and Developmental Biology* (Vol. 98, pp. 129–138). Elsevier Ltd. <https://doi.org/10.1016/j.semcdb.2019.05.029>
- Fiesel, F. C., Fričová, D., Hayes, C. S., Coban, M. A., Hudec, R., Bredenberg, J. M., Broadway, B. J., Markham, B. N., Yan, T., Boneski, P. K., Fiorino, G., Watzlawik, J. O., Hou, X., McCarty, A. M., Lewis-Tuffin, L. J., Zhong, J., Madden, B. J., Ordureau, A., An, H., ... Springer, W. (2023). Substitution of PINK1 Gly411 modulates substrate receptivity and turnover. *Autophagy*, 19(6), 1711–1732. <https://doi.org/10.1080/15548627.2022.2151294>
- Fujiwara, M., Marusawa, H., Wang, H. Q., Iwai, A., Ikeuchi, K., Imai, Y., Kataoka, A., Nukina, N., Takahashi, R., & Chiba, T. (2008). Parkin as a tumor suppressor gene for hepatocellular carcinoma. *Oncogene*, 27(46), 6002–6011. <https://doi.org/10.1038/onc.2008.199>
- Greene, N. P., Lee, D. E., Brown, J. L., Rosa, M. E., Brown, L. A., Perry, R. A., Henry, J. N., & Washington, T. A. (2015). Mitochondrial quality control, promoted by PGC-1 $\alpha$ , is dysregulated by Western diet-induced obesity and partially restored by moderate physical activity in mice. *Physiological Reports*, 3(7). <https://doi.org/10.14814/phy2.12470>
- Han, K., Hassanzadeh, S., Singh, K., Menazza, S., Nguyen, T. T., Stevens, M. V., Nguyen, A., San, H., Anderson, S. A., Lin, Y., Zou, J., Murphy, E., & Sack, M. N. (2017). Parkin regulation of CHOP modulates susceptibility to cardiac endoplasmic reticulum stress. *Scientific Reports*, 7(1). <https://doi.org/10.1038/s41598-017-02339-2>

- Hanna, R. A., Quinsay, M. N., Orogo, A. M., Giang, K., Rikka, S., & Gustafsson, Å. B. (2012). Microtubule-associated protein 1 light chain 3 (LC3) interacts with Bnip3 protein to selectively remove endoplasmic reticulum and mitochondria via autophagy. *Journal of Biological Chemistry*, 287(23), 19094–19104. <https://doi.org/10.1074/jbc.M111.322933>
- Ikeda, Y., Shirakabe, A., Maejima, Y., Zhai, P., Sciarretta, S., Toli, J., Nomura, M., Mihara, K., Egashira, K., Ohishi, M., Abdellatif, M., & Sadoshima, J. (2015). Endogenous Drp1 mediates mitochondrial autophagy and protects the heart against energy stress. *Circulation Research*, 116(2), 264–278. <https://doi.org/10.1161/CIRCRESAHA.116.303356>
- Jung, J., Zhang, Y., Celiku, O., Zhang, W., Song, H., Williams, B. J., Giles, A. J., Rich, J. N., Abounader, R., Gilbert, M. R., & Park, D. M. (2019). Mitochondrial Nix promotes tumor survival in the hypoxic niche of glioblastoma. *Cancer Research*, 79(20), 5218–5232. <https://doi.org/10.1158/0008-5472.CAN-19-0198>
- Khaminets, A., Behl, C., & Dikic, I. (2016). Ubiquitin-Dependent And Independent Signals In Selective Autophagy. In *Trends in Cell Biology* (Vol. 26, Issue 1, pp. 6–16). Elsevier Ltd. <https://doi.org/10.1016/j.tcb.2015.08.010>
- Kırsadere, İhsan, Dönmez, N., & Dönmez, H. H. (2019). The effects of quercetin on antioxidant and cytokine levels in rat hippocampus exposed to acute cadmium toxicity. *Journal of Cellular Neuroscience and Oxidative Stress*, 10(10), 10.
- Kırsadere, İ., Faruk Aydın, M., Usta, M., & Donmez, N. (n.d.). *Protective effects of oral melatonin against cadmium-*

*induced neurotoxicity in Wistar rats.*  
<https://doi.org/10.2478/aiht-2021-72-3513>

- Kubli, D. A., Zhang, X., Lee, Y., Hanna, R. A., Quinsay, M. N., Nguyen, C. K., Jimenez, R., Petrosyans, S., Murphy, A. N., & Gustafsson, Å. B. (2013). Parkin protein deficiency exacerbates cardiac injury and reduces survival following myocardial infarction. *Journal of Biological Chemistry*, 288(2), 915–926. <https://doi.org/10.1074/jbc.M112.411363>
- Landes, T., Emorine, L. J., Courilleau, D., Rojo, M., Belenguer, P., & Arnauné-Pelloquin, L. (2010). The BH3-only Bnip3 binds to the dynamin Opal to promote mitochondrial fragmentation and apoptosis by distinct mechanisms. *EMBO Reports*, 11(6), 459–465. <https://doi.org/10.1038/embor.2010.50>
- Li, X. C., Hu, Y., Wang, Z. H., Luo, Y., Zhang, Y., Liu, X. P., Feng, Q., Wang, Q., Ye, K., Liu, G. P., & Wang, J. Z. (2016). Human wild-type full-length tau accumulation disrupts mitochondrial dynamics and the functions via increasing mitofusins. *Scientific Reports*, 6. <https://doi.org/10.1038/srep24756>
- Liu, L., Feng, D., Chen, G., Chen, M., Zheng, Q., Song, P., Ma, Q., Zhu, C., Wang, R., Qi, W., Huang, L., Xue, P., Li, B., Wang, X., Jin, H., Wang, J., Yang, F., Liu, P., Zhu, Y., ... Chen, Q. (2012). Mitochondrial outer-membrane protein FUNDC1 mediates hypoxia-induced mitophagy in mammalian cells. *Nature Cell Biology*, 14(2), 177–185. <https://doi.org/10.1038/ncb2422>
- Lu, Y., Li, Z., Zhang, S., Zhang, T., Liu, Y., & Zhang, L. (2023). Cellular mitophagy: Mechanism, roles in diseases and small molecule pharmacological regulation.

*Theranostics*, 13(2), 736–766.  
<https://doi.org/10.7150/thno.79876>

- Montava-Garriga, L., & Ganley, I. G. (2020). Outstanding Questions in Mitophagy: What We Do and Do Not Know. In *Journal of Molecular Biology* (Vol. 432, Issue 1, pp. 206–230). Academic Press.  
<https://doi.org/10.1016/j.jmb.2019.06.032>
- Nunnari, J., & Suomalainen, A. (2012). Mitochondria: In sickness and in health. In *Cell* (Vol. 148, Issue 6, pp. 1145–1159). Elsevier B.V.  
<https://doi.org/10.1016/j.cell.2012.02.035>
- Özsan, M. (2023). Mitokondriyal Disfonksiyon ve Stres . In *Sağlık Bilimlerinde Öncü ve Çağdaş Çalışmalar: Vol. 33. Bölüm* (pp. 681–700). Duvar Yayınları.
- Özsan, M., & Ceylan, C. (2023). *Mitochondrial Dysfunction and Diabetes*. . 404–407.
- Özsan, M., Saygılı Düzova, Ü., & Dönmez, N. (2025). Neuroprotective role of curcumin on the hippocampus against the oxidative stress and inflammation of streptozotocin-induced diabetes in rats. *Metabolic Brain Disease*, 40(1), 24. <https://doi.org/10.1007/s11011-024-01438-0>
- Palikaras, K., Daskalaki, I., Markaki, M., & Tavernarakis, N. (2017). Mitophagy and age-related pathologies: Development of new therapeutics by targeting mitochondrial turnover. *Pharmacology & Therapeutics*, 178, 157–174.  
<https://doi.org/10.1016/J.PHARMTHERA.2017.04.005>
- Panigrahi, D. P., Praharaj, P. P., Bhol, C. S., Mahapatra, K. K., Patra, S., Behera, B. P., Mishra, S. R., & Bhutia, S. K. (2020a). The emerging, multifaceted role of mitophagy

- in cancer and cancer therapeutics. *Seminars in Cancer Biology*, 66, 45–58. <https://doi.org/10.1016/J.SEMCANCER.2019.07.015>
- Panigrahi, D. P., Praharaj, P. P., Bhol, C. S., Mahapatra, K. K., Patra, S., Behera, B. P., Mishra, S. R., & Bhutia, S. K. (2020b). The emerging, multifaceted role of mitophagy in cancer and cancer therapeutics. In *Seminars in Cancer Biology* (Vol. 66, pp. 45–58). Academic Press. <https://doi.org/10.1016/j.semcancer.2019.07.015>
- Panigrahi, D. P., Praharaj, P. P., Bhol, C. S., Mahapatra, K. K., Patra, S., Behera, B. P., Mishra, S. R., & Bhutia, S. K. (2020c). The emerging, multifaceted role of mitophagy in cancer and cancer therapeutics. In *Seminars in Cancer Biology* (Vol. 66, pp. 45–58). Academic Press. <https://doi.org/10.1016/j.semcancer.2019.07.015>
- Pickrell, A. M., & Youle, R. J. (2015). The roles of PINK1, Parkin, and mitochondrial fidelity in parkinson's disease. In *Neuron* (Vol. 85, Issue 2, pp. 257–273). Cell Press. <https://doi.org/10.1016/j.neuron.2014.12.007>
- Pryde, K. R., Smith, H. L., Chau, K. Y., & Schapira, A. H. V. (2016). PINK1 disables the anti-fission machinery to segregate damaged mitochondria for mitophagy. *Journal of Cell Biology*, 213(2), 163–171. <https://doi.org/10.1083/jcb.201509003>
- Rovira-Llopis, S., Bañuls, C., Diaz-Morales, N., Hernandez-Mijares, A., Rocha, M., & Victor, V. M. (2017). Mitochondrial dynamics in type 2 diabetes: Pathophysiological implications. In *Redox Biology* (Vol. 11, pp. 637–645). Elsevier B.V. <https://doi.org/10.1016/j.redox.2017.01.013>

- Rugarli, E. I., & Langer, T. (2012). Mitochondrial quality control: A matter of life and death for neurons. *EMBO Journal*, 31(6), 1336–1349. <https://doi.org/10.1038/emboj.2012.38>
- Scheele, C., Nielsen, A. R., Walden, T. B., Sewell, D. A., Fischer, C. P., Brogan, R. J., Petrovic, N., Larsson, O., Tesch, P. A., Wennmalm, K., Hutchinson, D. S., Cannon, B., Wahlestedt, C., Pedersen, B. K., & Timmons, J. A. (2007). Altered regulation of the PINK1 locus: a link between type 2 diabetes and neurodegeneration? *The FASEB Journal*, 21(13), 3653–3665. <https://doi.org/10.1096/fj.07-8520com>
- Sciarretta, S., Maejima, Y., Zablocki, D., & Sadoshima, J. (2018). The Role of Autophagy in the Heart. *The Annual Review of Physiology Is Online at Downloaded from Www.Annualreviews.Org. Guest (Guest, 39, 5.* <https://doi.org/10.1146/annurev-physiol-021317>
- Sekine, S., & Youle, R. J. (2018). PINK1 import regulation; a fine system to convey mitochondrial stress to the cytosol. In *BMC Biology* (Vol. 16, Issue 1). BioMed Central Ltd. <https://doi.org/10.1186/s12915-017-0470-7>
- Shirihai, O. S., Song, M., & Dorn, G. W. (2015). How mitochondrial dynamism orchestrates mitophagy. In *Circulation Research* (Vol. 116, Issue 11, pp. 1835–1849). Lippincott Williams and Wilkins. <https://doi.org/10.1161/CIRCRESAHA.116.306374>
- Song, M., Zhao, X., & Song, F. (2021). Aging-Dependent Mitophagy Dysfunction in Alzheimer’s Disease. In *Molecular Neurobiology* (Vol. 58, Issue 5, pp. 2362–2378). Springer. <https://doi.org/10.1007/s12035-020-02248-y>

- Sowter, H. M., Ratcliffe, P. J., Watson, P., Greenberg, A. H., & Harris, A. L. (2001). HIF-1-dependent Regulation of Hypoxic Induction of the Cell Death Factors BNIP3 and NIX in Human Tumors 1. In *CANCER RESEARCH* (Vol. 61). <http://www.ncbi.nlm.nih.gov/SAGE>.
- Suen, D. F., Narendra, D. P., Tanaka, A., Manfredi, G., & Youle, R. J. (2010). Parkin overexpression selects against a deleterious mtDNA mutation in heteroplasmic hybrid cells. *Proceedings of the National Academy of Sciences of the United States of America*, 107(26), 11835–11840. <https://doi.org/10.1073/pnas.0914569107>
- Tanaka, A., Cleland, M. M., Xu, S., Narendra, D. P., Suen, D. F., Karbowski, M., & Youle, R. J. (2010). Proteasome and p97 mediate mitophagy and degradation of mitofusins induced by Parkin. *Journal of Cell Biology*, 191(7), 1367–1380. <https://doi.org/10.1083/jcb.201007013>
- Um, J. H., & Yun, J. (2017). Emerging role of mitophagy in human diseases and physiology. In *BMB Reports* (Vol. 50, Issue 6, pp. 299–307). The Biochemical Society of the Republic of Korea. <https://doi.org/10.5483/BMBRep.2017.50.6.056>
- Uoselis, L., Nguyen, T. N., & Lazarou, M. (2023). Mitochondrial degradation: Mitophagy and beyond. In *Molecular Cell* (Vol. 83, Issue 19, pp. 3404–3420). Cell Press. <https://doi.org/10.1016/j.molcel.2023.08.021>
- Vara-Perez, M., Felipe-Abrio, B., & Agostinis, P. (2019). Mitophagy in cancer: A tale of adaptation. In *Cells* (Vol. 8, Issue 5). MDPI. <https://doi.org/10.3390/cells8050493>
- Vives-Bauza, C., Zhou, C., Huang, Y., Cui, M., De Vries, R. L. A., Kim, J., May, J., Tocilescu, M. A., Liu, W., Ko, H. S., Magrané, J., Moore, D. J., Dawson, V. L., Grailhe,

- R., Dawson, T. M., Li, C., Tieu, K., & Przedborski, S. (2010). PINK1-dependent recruitment of Parkin to mitochondria in mitophagy. *Proceedings of the National Academy of Sciences of the United States of America*, *107*(1), 378–383. <https://doi.org/10.1073/pnas.0911187107>
- Whelan, R. S., Kaplinskiy, V., & Kitsis, R. N. (2010). Cell death in the pathogenesis of heart disease: Mechanisms and significance. In *Annual Review of Physiology* (Vol. 72, pp. 19–44). <https://doi.org/10.1146/annurev.physiol.010908.163111>
- Wong, Y. C., & Holzbaur, E. L. F. (2014). Optineurin is an autophagy receptor for damaged mitochondria in parkin-mediated mitophagy that is disrupted by an ALS-linked mutation. *Proceedings of the National Academy of Sciences of the United States of America*, *111*(42), E4439–E4448. <https://doi.org/10.1073/pnas.1405752111>
- Yamano, K., & Youle, R. J. (2013). PINK1 is degraded through the N-end rule pathway. *Autophagy*, *9*(11), 1758–1769. <https://doi.org/10.4161/auto.24633>
- Youle, R. J., & Narendra, D. P. (2011). Mechanisms of mitophagy. *Nature Reviews Molecular Cell Biology*, *12*(1), 9–14. <https://doi.org/10.1038/nrm3028>
- Youle, R. J., & Van Der Blik, A. M. (2012). Mitochondrial fission, fusion, and stress. In *Science* (Vol. 337, Issue 6098, pp. 1062–1065). American Association for the Advancement of Science. <https://doi.org/10.1126/science.1219855>
- Zhang, C., Lin, M., Wu, R., Wang, X., Yang, B., Levine, A. J., Hu, W., & Feng, Z. (2011). Parkin, a p53 target gene, mediates the role of p53 in glucose metabolism and the

Warburg effect. *Proceedings of the National Academy of Sciences of the United States of America*, 108(39), 16259–16264. <https://doi.org/10.1073/pnas.1113884108>

Zhang, J., & Ney, P. A. (2009). Role of BNIP3 and NIX in cell death, autophagy, and mitophagy. In *Cell Death and Differentiation* (Vol. 16, Issue 7, pp. 939–946). <https://doi.org/10.1038/cdd.2009.16>

Zhu, J., Wang, K. Z. Q., & Chu, C. T. (2013). After the banquet: Mitochondrial biogenesis, mitophagy, and cell survival. In *Autophagy* (Vol. 9, Issue 11, pp. 1663–1676). Taylor and Francis Inc. <https://doi.org/10.4161/auto.24135>